characteristics in Botswana

### Check for updates

Copyright © 2023 1... Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works

#### Roger L. Shapiro<sup>1,2</sup>\*, Gbolahan Ajibola<sup>2</sup>, Kenneth Maswabi<sup>2</sup>, Michael Hughes<sup>3</sup>, Bryan S. Nelson<sup>3</sup>, Aischa Niesar<sup>4</sup>, Molly Pretorius Holme<sup>1</sup>, Kathleen M. Powis<sup>1,2,5</sup>, Maureen Sakoi<sup>2</sup>, Oganne Batlang<sup>2</sup>, Sikhulile Moyo<sup>1,2</sup>, Terence Mohammed<sup>2</sup>, Comfort Maphorisa<sup>2</sup>, Kara Bennett<sup>6</sup>, Zixin Hu<sup>7</sup>, Francoise Giguel<sup>7</sup>, Jacqueline D. Reeves<sup>8</sup>, Michael A. Reeves<sup>8</sup>, Ce Gao<sup>4</sup>, Xu Yu<sup>4</sup>, Margaret E. Ackerman<sup>9</sup>, Adrian McDermott<sup>10</sup>, Marlene Cooper<sup>11</sup>, Marina Caskey<sup>12</sup>, Lucio Gama<sup>10</sup>, Patrick Jean-Philippe<sup>13</sup>, Dwight E. Yin<sup>13</sup>, Edmund V. Capparelli<sup>14</sup>, Shahin Lockman<sup>1,2,7</sup>, Joseph Makhema<sup>1,2</sup>, Daniel R. Kuritzkes<sup>7</sup>+, Mathias Lichterfeld<sup>4,7</sup>+

Broadly neutralizing antibody treatment maintained

HIV suppression in children with favorable reservoir

Broadly neutralizing antibodies (bNAbs) may provide an alternative to standard antiretroviral treatment (ART) for controlling HIV-1 replication and may have immunotherapeutic effects against HIV-1 reservoirs. We conducted a prospective clinical trial with two HIV-1 bNAbs (VRC01LS and 10-1074) in children (*n* = 25) who had previously initiated small-molecule ART treatment before 7 days of age and who continued treatment for at least 96 weeks. Both bNAbs were dosed intravenously every 4 weeks, overlapping with ART for at least 8 weeks and then continued for up to 24 weeks or until detectable viremia of HIV-1 RNA rose above 400 copies per milliliter in the absence of ART. Eleven (44%) children maintained HIV-1 RNA below 400 copies per milliliter through 24 weeks of bNAb-only treatment; 14 (56%) had detectable viremia above 400 copies per milliliter at a median of 4 weeks. Archived HIV-1 provirus susceptible to 10-1074, lower birth HIV-1 DNA reservoir in peripheral blood mononuclear cells, sustained viral suppression throughout early life, and combined negative qualitative HIV-1 DNA polymerase chain reaction and negative HIV-1 serology at entry were associated with maintaining suppression on bNAbs alone. This proof-of-concept study suggests that bNAbs may represent a promising treatment modality for infants and children living with HIV-1. Future studies using newer bNAb combinations with greater breadth and potency are warranted.

#### INTRODUCTION

HIV

Broadly neutralizing antibodies (bNAbs) against HIV-1 are an emerging treatment option for people living with HIV-1 with the potential to maintain HIV-1 RNA suppression (1, 2). bNAbs can be administered infrequently, which avoids adherence concerns of daily oral antiretroviral treatment (ART), may limit long-term toxicity from prolonged ART, and may enhance immune responses and deplete residual viral reservoirs, offering a potential pathway to post-treatment viral control in some individuals (3, 4). Children living with HIV-1 who have been treated continuously from birth are an ideal group for bNAb treatment, because they have limited viral reservoirs (5) and may be less likely to have preexisting viral

\*Corresponding author. Email: rshapiro@hsph.harvard.edu †These authors contributed equally to this work. Downloaded from https://www.science.org at Harvard University on July 05 ces for adheris some similar se 1/2, use of

resistance to bNAbs (6). Children are also ideal candidates for ART-sparing strategies that avoid long-term toxicities and adherence considerations with daily dosing.

Long-term HIV-1 RNA suppression has been reported in some adult trials using bNAbs in combination (3, 7), warranting similar studies in pediatric populations. The Tatelo study was a phase 1/2, single-arm, multisite clinical trial to evaluate the combined use of two bNAbs, VRC01LS and 10-1074. VRC01 targets the CD4 binding site, and the LS formulation has a modified Fc receptor that increases its half-life. VRC01 had been used in a pediatric trial previously (8), and a pediatric study of VRC01LS was underway at the time of starting Tatelo (9). 10-1074 targets the V3 loop and has been used as combined treatment with CD4 binding agents in previous adult (2) (but not pediatric) studies. The Tatelo study administered both of these bNAbs as monthly treatments in a cohort of children in Botswana who had received ART since birth in a previous study (5, 10).

#### RESULTS

#### Participant characteristics

Between March 2020 and January 2021, all 28 (70%) Early Infant Treatment (EIT) study participants who were eligible for the Tatelo study were enrolled (Fig. 1); the remaining 12 (30%) EIT study participants were ineligible for Tatelo because of detectable viremia of HIV-1 RNA of  $\geq$ 40 copies/ml within the preceding 24 weeks. Of the 28 Tatelo enrollees, 14 (50%) had never experienced

<sup>&</sup>lt;sup>1</sup>Department of Immunology and Infectious Diseases, Harvard T. H. Chan School of Public Health, Boston, MA 02115, USA. <sup>2</sup>Botswana Harvard Health Partnership, Gaborone, Botswana. <sup>3</sup>Department of Biostatistics, Harvard T. H. Chan School of Public Health, Boston, MA 02115, USA. <sup>4</sup>Ragon Institute of MGH, MIT, and Harvard, Cambridge, MA 02139, USA. <sup>5</sup>Departments of Internal Medicine and Pediatrics, Massachusetts General Hospital, Boston, MA 02114, USA. <sup>6</sup>Bennett Statistical Consulting Inc., Ballston Lake, NY 12019, USA. <sup>7</sup>Division of Infectious Diseases, Brigham and Women's Hospital, Boston, MA 02115, USA. <sup>8</sup>Labcorp-Monogram Biosciences Inc., South San Francisco, CA 94080, USA. <sup>9</sup>Thayer School of Engineering, Dartmouth College, Hanover, NH 03755, USA. <sup>10</sup>Vaccine Research Center, Bethesda, MD 20892, USA. <sup>11</sup>Frontier Science & Technology Research Foundation Inc., Amherst, NY 14226, USA. <sup>12</sup>Rockefeller University, New York, NY 10065, USA. <sup>13</sup>National Institute of Allergy and Infectious Diseases, National Institutes of Health, Rockville, MD 20892, USA. <sup>14</sup>Department of Pediatrics, University of California San Diego, La Jolla, CA 92037, USA.



Fig. 1. Flow diagram. N = 40 children in the Early Infant Treatment (EIT) study were potentially eligible for inclusion in Tatelo; 28 enrolled in the ART + bNAb overlap step, and 25 continued into the bNAb-only step.

detectable viremia of  $\geq$ 40 copies/ml during EIT participation, and 14 (50%) had experienced viremia of  $\geq$ 40 copies/ml at least once. Enrolled children had a median age of 3.6 (range: 2.4 to 5.6) years, 19 (68%) were female, and the median entry CD4 T cell count was 1198 with an interquartile range (IQR) of 843 to 1684 cells/mm<sup>3</sup> (Table 1). All enrolled children were screened for HIV-1 RNA before entry into the ART and bNAbs step. HIV-1 RNA was >40 copies/ml in two children on the day of bNAb initiation and in one child after 4 weeks of ART/bNAbs; thus, bNAb treatment was discontinued in these three children, ART was continued, and adherence was reinforced, leading to subsequent viral resuppression.

#### Virologic and immunologic outcomes

Twenty-five (89%) children completed the ART plus bNAb step (six for 32 weeks and 19 for 8 weeks) and advanced to the bNAb-only step of the study. Of these, 11 children [44%, 95% confidence interval (CI): 24 to 65%] maintained an HIV-1 RNA of <400 copies/ml (defined as bNAb successes), and 10 children (40%, 95% CI: 21 to 61%) maintained <40 copies/ml for 24 weeks (Fig. 2A); one child had a single HIV-1 RNA value of 234 copies/ml at week 16, with <40 copies/ml at all other weeks. Fourteen children (56%) had detectable viremia of  $\geq$ 400 copies/ml at a median of 4 (range: 1 to 20) weeks (defined as bNAb failures) and were restarted on ART at a

Shapiro et al., Sci. Transl. Med. 15, eadh0004 (2023) 5 July 2023

median of 4 (range: 1 to 7) days from first detected viremia. Kaplan-Meier estimates for time to first HIV-1 RNA of  $\geq$ 400 copies/ml are shown in Fig. 2B. Among children with failure, median HIV-1 RNA at ART restart was 4.42 (range: 2.87 to 6.42) log<sub>10</sub> copies/ml. After failure, all children had viral resuppression to <40 copies/ml on their previous ART regimen [lopinavir/ritona-vir (LPV/r)–based in all cases], at a median of 4 (range: 1 to 20) weeks from ART restart. CD4 T cell counts in children who experienced detectable viremia of  $\geq$ 400 copies/ml were similar to post-intervention CD4 cell counts in children who did not, with a median above 1000 copies/mm<sup>3</sup> in both groups. No child in either group had a concerning pattern of CD4 T cell count decline during the study (fig. S1).

#### Safety, pharmacokinetics, and antidrug antibody analysis

No infusion reactions occurred, and bNAbs were well tolerated. Three children experienced a total of five grade 3 events, with one (neutropenia) during the bNAb/ART step considered possibly related to bNAb treatment (tables S1 and S2). There were no grade 4 events. Pharmacokinetic (PK) troughs before each dose in the bNAb-only step revealed adequate 10-1074 and VRC01LS concentrations for all children. Overall predose troughs were in the expected range and were consistently above 100 µg/ml for both 
 Table 1. Baseline characteristics of enrolled Tatelo participants (N =

 28). IQR, interquartile range.

#### **Birth characteristics**

Female sex, no. (%)	19 (68)
Gestational age at birth*, no. (%)	
35 weeks	4 (14)
36 weeks	6 (21)
37 weeks	2 (7)
38–41 weeks	16 (57)
Median birth weight, kg (IQR)	2.95 (2.60–3.20)
Median HIV-1 RNA at birth, log <sub>10</sub> copies/ml (IQR)	4.09 (2.54–4.65)
Median HIV DNA at birth, copies per million cells (IQR) <sup>+</sup>	490 (121–1221)
Maternal education, highest level attained, no. (%)	
None/primary	6 (21)
Junior secondary	13 (46)
Senior secondary	4 (14)
Tertiary	5 (18)
Maternal employment status, no. (%)	
Salaried	6 (21)
Paid domestic work	2 (7)
None/unemployed	20 (71)
Characteristics at time of enrollment to Tatelo	
Median age, years (IQR)	3.60 (3.10–4.46)
Median weight, kg (IQR)	12.90 (11.90–15.20)
ART regimen‡ – no.	
ZDV + 3TC + LPV/r	26
ABC + 3TC + LPV/r	1
ZDV + ABC + 3TC + LPV/r	1
Median CD4 count, cells/mm <sup>3</sup> (IQR)	1198 (843–1684)
Median HIV DNA (at 84/96 weeks of age), copies per million cells (IQR)	35.3 (8.38–102.3)
*EIT study oveluded children hern hefere 25 weeks gest	ational

\*Ell study excluded children born before 35 weeks gestational age. †Measured in PBMCs by ddPCR. ‡ZDV, zidovudine; 3TC, lamivudine; LPV/r, lopinavir/ritonavir; ABC, abacavir.

bNAbs: median of 211.0  $\mu$ g/ml (IQR: 183.6 to 259.2) for 10-1074 and 259.6  $\mu$ g/ml (IQR: 201.0 to 305.6) for VRC01LS. Although fewer values were available for those with bNAb failure who exited the study step early, median trough bNAb values were similar for successes and failures during the bNAb-only step (fig. S2). No antidrug antibodies were observed to 10-1074 or VRC01LS.

#### Characteristics of children with successful bNAb treatment

Children with bNAb-only treatment success had favorable preintervention clinical and viral reservoir characteristics (Table 2). Almost all children who succeeded (nine, 82%) had sustained viral suppression (<40 copies/ml) at all EIT visits from initial suppression through Tatelo entry, compared with 29% of children who failed. In addition, the total HIV DNA in peripheral blood mononuclear cells (PBMCs) near birth was significantly lower in children who succeeded (P = 0.02). Most children who succeeded had negative qualitative DNA (nine, 82%) or negative enzyme immunoassay (EIA; nine, 82%), or both (eight, 73%), at Tatelo entry; in contrast, none of the children who failed had a "negative/negative" pattern at Tatelo entry (73 versus 0%, P < 0.001).

#### Neutralization and reservoir quantification assays

Neutralization assay data were limited either by amplification failure or failure of the cloning or cell assay step. In all plasma samples with a successful assay at time of detectable viremia of  $\geq$ 400 copies/ml (bNAb failure), some degree of reduced neutralization by 10-1074 or VRC01LS was observed (Fig. 2A). Viral envelope sequences from intact proviruses collected near birth had a largely similar pattern as plasma at bNAb failure when matched results were available. Among virologic successes on bNAbs, archived provirus demonstrated dual bNAb susceptibility for three (50%) of six with available results, and all had complete (83%) or partial (17%) 10-1074 susceptibility. In contrast, only two (29%) of seven failures with available results had complete or partial susceptibility to 10-1074 in archived provirus (P = 0.02).

Digital droplet polymerase chain reaction (ddPCR) and near full-length proviral sequencing were performed at study entry and several follow-up time points, but most children had values below the limit of detection (LOD; likely due to low viral reservoirs and the limited number of PBMCs that could be safely collected from children). Birth ddPCR in PBMCs predicted successful dual bNAb treatment (Table 2), and a similar pattern was observed for full-length individual proviral sequencing (FLIP-seq) (Fig. 3). At entry to Tatelo, both defective provirus and total provirus DNA (intact plus defective) by FLIP-seq were more commonly above the LOD in children who later failed (each 79%) than those who succeeded (each 18%; P = 0.005). Among failures, detectable intact provirus was observed in three individuals (21%) at entry and seven individuals (50%) after failure. These findings were not attributable to variations in cell numbers used for measurements (fig. S3).

#### DISCUSSION

As the breadth, potency, and half-lives of bNAbs improve, immunebased HIV treatment may offer advantages for children facing lifelong ART. We found that monthly VRC01LS and 10-1074 dual bNAb infusions were well tolerated and maintained a viral suppression of <400 copies/ml for 24 weeks in 44% of children who had been treated with ART from birth. This proof-of-concept study provides early evidence that current bNAb combinations may offer an alternative to small-molecule ART in selected children with favorable clinical and resistance characteristics and that future longacting bNAb combinations may expand the benefits of this strategy to more children living with HIV.

Our results were generally concordant with adult treatment studies using similar bNAb combinations, although virologic success was somewhat lower in our study. Among adults with chronic HIV subtype B, treatment with 10-1074 plus 3BNC117 maintained viral suppression in 76% of participants for 20 weeks after ART discontinuation (3). This same combination maintained viral suppression for all adults with prescreened susceptible virus in two separate studies (7, 11). Plasma prescreening for HIV-1 susceptibility to bNAbs in our study was not possible for our virally suppressed cohort, but our data suggest that screening archived



**Fig. 2. Treatment outcomes in the Tatelo study.** (**A**) Shown on the left are available participant antibody neutralization assay results for *env* amplicons of full-length intact provirus near birth and plasma at failure. Proviral samples from PBMCs at baseline were available for all participants, but amplification succeeded in only 14 of 25. Baseline amplicons were from birth (85%), 4 to 24 weeks (13%), or 84 weeks (2%). Plasma samples were available for 14 failures but amplified in only 8. We defined full susceptibility to each bNAb as 90% inhibitory concentration ( $IC_{90}$ ) of  $\leq 1.0 \mu g/ml$  and maximum percent inhibition (MPI) of  $\geq 98\%$ . The plot on the right shows bNAb-only step HIV-1 RNA outcomes, grouped by failures (top, orange) and successes (bottom, blue). Participant HIV-1 RNA outcomes are shown by bNAb-only week; the bars extend through week of completion of this study step. Values at ends of bars indicate HIV-1 RNA copies/ml at first virologic failure. Each row in (A) indicates the same participant. (**B**) Shown is the cumulative proportion of participants with HIV-1 RNA-detectable viremia of  $\geq 400$  copies/ml over time during the bNAb-only phase. The shaded area shows the 95% CI.

provirus stored from pre-ART birth samples might identify the children who are most likely to maintain viral suppression on bNAbs. Neutralization assay results in our study were limited because of assay failure or inability to amplify full-length intact proviral DNA (in part due to very low viral reservoirs in the cohort), but a pattern of reduced susceptibility to one or both bNAbs was observed for all children with results at the time of virologic failure. This same pattern was also present in archived provirus when matching samples were available and in all but one when only archived provirus was available. In contrast, we found that, for half of children who succeeded, archived proviruses were susceptible to both bNAbs; furthermore, all analyzed proviruses from successes had at least partial susceptibility to 10-1074. Prescreening may be particularly important for regions where subtype C predominates (including Botswana), because there is less inherent susceptibility to most current bNAbs in these regions (*12*).

Our study also advances the concept of prescreening for future pediatric cohorts beyond the use of neutralization assays, because

Baseline/enrollment characteristics	Total ( <i>N</i> = 25)*	Treatment success on bNAbs (N = 11) Median (IQR) or numb	Treatment failure on bNAbs (N = 14) er (%)	P value
Age at ART start (days)	3 (2–4)	3 (3–5)	2 (2–3)	0.10
Age at bNAb start (years)	3.70 (3.10–4.40)	4.20 (3.40–4.60)	3.45 (2.90–4.40)	0.26
HIV-1 RNA at birth (copies/ml)	3145 (310–25,507)	2279 (381–12,984)	20,465 (292–33,502)	0.37
HIV-1 RNA undetectable since 24 weeks <sup>+</sup>				
Yes	13 (52%)	9 (82%)	4 (29%)	0.02
No	12 (48%)	2 (18%)	10 (71%)	
Total HIV DNA in PBMCs at birth by ddPCR (copies per $10^6$ )‡	465 (100–1129)	155 (46–465)	784 (166–1246)	0.02
Intact HIV DNA in PBMCs at birth by FLIP-seq (copies per 10 <sup>6</sup> ) <sup>‡</sup>	2.9 (0.22–280.4)	1.16 (0.22–38.3)	4.59 (0.52–280.4)	-§
Number with intact HIV DNA in PBMCs at birth by FLIP-seq (for those >LOD)	18 (72%)	6 (55%)	12 (86%)	0.18
Intact HIV DNA in PBMCs at Tatelo entry by FLIP-seq (copies per 10 <sup>6</sup> ) <sup>‡</sup>	0.24 (0.04–2)	0.26 (0.04–2)	0.22 (0.08–1)	-§
Number with intact HIV DNA in PBMCs at Tatelo entry by FLIP- seq (for those >LOD)	5 (20%)	2 (18%)	3 (21%)	>0.99
Negative whole-blood qualitative HIV DNA PCR and HIV enzym	e immunoassay a	t Tatelo entry		
Yes	8 (32%)	8 (73%)	0	<0.001
No	17 (68%)	3 (27%)	14 (100%)	
Amount of bNAb/ART overlap				
32 weeks	6 (24%)	5 (45%)	1 (7%)	0.06
8 weeks	19 (76%)	6 (55%)	13 (93%)	
CD4 T cell count (cells/mm <sup>3</sup> )				
Start of bNAb-only treatment	1149 (922–1502)	984 (808–1185)	1380 (1004–1868)	0.05
bNAb susceptibility				
10-1074-susceptible (PBMCs at birth)	8 (57%)	6 (100%)	2 (25%)	0.01
VRC01LS-susceptible (PBMCs at birth)	7 (50%)	3 (50%)	4 (50%)	>0.99

#### Table 2. Characteristics by response group for bNAb-only treatment with VRC01LS and 10-1074 in children living with HIV in Botswana.

\*Excludes three children who began bNAbs but never discontinued ART. \*Defined as all visits at or after 24 weeks of age with HIV-1 RNA of <40 copies/ml; per protocol, all HIV-1 RNA values in the 24 weeks before bNAb initiation must be <40 copies/ml. \*An imputed value of one provirus in double the number of analyzed cells used, if no target identification. SLow sample size and high proportion below the LOD precluded comparison of distributions.  $\|N = 14$ (eight treatment failures and six treatment successes) with amplification. bNAb susceptibility was based on a maximum percent inhibition (MPI) of  $\geq$ 98%.

we identified several additional markers for virologic success with bNAb treatment. HIV DNA in PBMCs from birth was significantly lower in those who succeeded on bNAbs alone (P = 0.02), suggesting a potential advantage in some children from very early life. Most of the children who succeeded also had sustained viral suppression before the bNAb intervention, whereas the majority of those who failed had experienced detectable viremia of  $\geq$ 40 copies/ml at some point in early life. Viremia before bNAb intervention may have therefore led to some bNAb resistance, and its detection served as a useful marker for bNAb failure. Last, an additional predictor of success was the combination of having reverted to a negative whole-blood HIV qualitative DNA test at Tatelo entry and

never developing positive serology by EIA; all eight children with this pattern succeeded. These simple clinical markers are widely available, and we believe that this combination of negative/negative should be explored as a real-time biomarker for entry into future pediatric bNAb treatment studies. Of the three children who succeeded without the negative/negative pattern, two had detectable intact provirus integrated into nonencoding regions of the genome, and this "locked" pattern is being further studied (13); the third was one of a small number of successes with several episodes of detectable viremia before Tatelo and had a positive EIA at entry. Note that each of the characteristics for success listed above were closely interrelated, and because of small numbers,



**Fig. 3. Differences in intact, defective, and total HIV-1 by FLIP-seq at birth, 84 to 96 weeks, Tatelo study entry, and end of bNAb treatment among successes and failures.** (A to C) Shown is the quantification of the viral reservoir obtained by FLIP-seq. Intact (A), defective (B), and total (C) proviruses per 1 million PBMCs were identified at birth, week 84 to 96 of age, Tatelo entry, and end of bNAb treatment. Study participants were divided into two groups according to success or failure to maintain viral control (HIV-1 RNA of <400 copies/ml) throughout the bNAb-only step. Open symbols represent the LOD when no virus was detectable, with an imputed value of 1 provirus in double the number of analyzed cells without target identification. The range of the number of PBMCs across all time points and both groups was  $1.51 \times 10^6$  to  $1.11 \times 10^7$  cells (median =  $1.45 \times 10^6$ ); for those below LOD, the range of the number of PBMCs was  $6.06 \times 10^4$  to  $4.31 \times 10^6$  cells (median =  $1.0 \times 10^6$ ).

multivariable analysis was not possible. Likewise, although five of six children who received 32 weeks of ART/bNAb overlap succeeded, these children all had additional favorable characteristics, limiting our ability to assess whether the longer overlap period improved outcomes.

Whether bNAbs were directly responsible for maintaining viral suppression in our study or whether some children would have maintained viral suppression without ART or bNAbs remains an open question. Using data from the Children with HIV Early Antiretroviral Therapy (CHER) study (14, 15), we prespecified that a 24-week success of  $\geq$ 30% would be unlikely to occur by chance, even among low-reservoir children. The neutralization data provided additional support for a causal role of the bNAbs but were not

definitive because of small numbers and because we cannot exclude the possibility that higher bNAb sensitivity tracked with more limited viral diversity. A randomized design comparing bNAbs with an analytic treatment interruption (ATI) was not considered feasible at the time Tatelo was conducted, but for children with low or undetectable reservoirs and a constellation of favorable markers, an ATI component could be considered in future trials. Given the potential vaccinal effect (16, 17) and reservoir-lowering effect (1, 4, 18, 19) previously described for bNAbs, candidates for such a trial may benefit from a period of bNAbs before ATI to maximize the chance for success.

There were no safety concerns raised for VRC01LS or 10-1074 in our study, and both were well tolerated, as expected from previous studies (2, 19-21). Few grade 3 (and no grade 4) events were reported, and none resulted in bNAb discontinuation. CD4 T cell counts were largely unaffected between the beginning and end of the study. Trough PK values were sufficient at nearly all time points and did not differ between successes and failures. The slow elimination of VRC01LS and frequent (every 4 weeks) administration resulted in trough concentrations substantially higher than previous VRC01 therapy switch (20) and preexposure prophylaxis studies (22). Monthly bNAb infusions were also highly acceptable to caregivers of the participants in the study (23).

Limitations of our study were the small sample size, including the limited ability to perform neutralization assays and to quantify changes in intact viral reservoir over time, both of which were unavoidable challenges related to the extraordinarily low reservoirs in this cohort. Children treated with ART from birth and virally suppressed at bNAb initiation may not be representative of other children with HIV. All study participants are now in long-term followup, and in-depth profiling of reservoir cells and immune responses is ongoing. Our study benefitted from having complete treatment and reservoir data on a cohort of children followed from birth, from no losses to follow-up, and from our ability to ensure that ART could be safely restarted when needed. In conclusion, nearly half of children who had received ART from birth maintained viral suppression for 24 weeks with monthly 10-1074 and VRC01LS alone, and easily identifiable markers predicted successful outcomes. These findings support the use of bNAbs with greater breadth and potency in future pediatric trials and provide a methodology to easily screen participants with the greatest chance for success.

#### **MATERIALS AND METHODS**

#### Study design

The Tatelo study evaluated dual bNAbs as a treatment alternative in children. The study began with an intensive PK and safety phase for 10-1074 and VRC01LS whileART was continued-first individually among 12 participants (six per bNAb) and then during dual administration among six of the first 12 participants. Results were previously reported from this intensive PK and safety evaluation (24). The main study consisted of a single arm; participants in the main study were followed through three steps. The first was an ART/bNAb overlap step, where ART was continued while dual bNAbs were administered every 4 weeks. This step lasted 32 weeks for the first six participants (for planned PK and safety analyses and Safety Monitoring Committee approval before permitting additional enrollment in the step) and 8 weeks for subsequent participants. The next step was the bNAb-only step, where ART was discontinued for up to 24 weeks (or until any detectable viremia of  $\geq$ 400 copies/ml). At 24 weeks (or upon detectable viremia of ≥400 copies/ml), participants discontinued bNAbs and resumed ART and were followed for an additional 24 weeks in the ART restart step. ART regimens at study entry and upon restart were per Botswana guidelines and consisted of LPV/r-based three-drug oral ART in all children.

#### Trial ethics and oversight

The study was approved by Institutional Review Boards in Botswana (Human Research Development Committee, HPDME 13/18/1 X1) and at the Harvard T. H. Chan School of Public Health (Harvard Longwood Campus Institutional Review Board, IRB18-0062). A parent or guardian provided written informed consent for all participants. The study was monitored by an independent Safety Monitoring Committee.

#### Study population and monitoring

All Tatelo participants had previously taken part in the EIT study, a clinical trial evaluating early infant HIV-1 diagnosis and treatment in Gaborone and Francistown, Botswana (NCT02369406) (5, 10). Children eligible for Tatelo were EIT participants treated since before 7 days of age, on continuous ART for  $\geq$ 96 weeks, and with an HIV-1 RNA of <40 copies/ml for at least 24 weeks before entry. Tatelo visits occurred at least every 4 weeks during bNAb administration and at 1- to 2-week intervals during the bNAb-only step. All adverse events (grade 1 or higher) (25), including any infusion reactions, were recorded.

#### Laboratory testing

HIV-1 RNA (Abbott m2000sp/m2000rt, Abbott Molecular Inc.) quantified to a threshold of 40 copies/ml and qualitative DNA PCR (Roche Cobas AmpliPrep/Cobas TaqMan HIV-1 Qualitative PCR) testing were performed at the Botswana Harvard HIV Reference Laboratory, Gaborone, Botswana every 1 to 2 weeks during the bNAb-only step. Safety and monitoring evaluations included hematology, serum chemistry, CD4 and CD8 T cell count, and EIA serologic HIV testing at 4- to 8-week intervals during bNAb administration. EIA was performed in parallel using Murex HIV-1.2.O (DiaSorin), Bio-Rad Genetic Systems HIV-1/HIV-2 Plus O, or Abbott ARCHITECT i1000SR (Abbott Diagnostics). Trough PK testing occurred before each bNAb dose, and antidrug antibody (ADA) testing occurred at entry and after final bNAb dosing. bNAb PK testing was performed as described (24). ADA testing was per-formed by three-tiered approach for VRC01LS at the Vaccine Re-search Center (21, 26) and by electrochemiluminescence bridging assay for 10-1074 at Dartmouth College (27, 28). PBMCs were collected at least monthly during bNAb steps. ddPCR testing for a subset of specimens and FLIP-seq (29, 30) at entry and at last bNAb receipt were performed at the Ragon Insti-tute. ddPCR was performed using the QX100 Droplet Digital PCR System (Bio-Rad) using primers and probes described previously

System (Bio-Rad) using primers and probes described previously (31) (127-base pair 5'-LTR-gag amplicon; coordinates 684 to 810 in HIV-1 reference strain HXB2) and normalized to the RPP30 gene. When viral copies were undetectable, data were reported as "LOD" (calculated as 0.2 copies per maximum number of cells tested without target identification) (5). On the basis of ddPCR results and Poisson distribution statistics, genomic DNA was diluted to single HIV-1 copies and subjected to HIV-1 near-full genome amplification using a one amplicon approach with primers described previously (5). When there were no HIV-1 amplification products detectable, results were reported as LOD and calculated as 0.5 copies per maximum number of cells analyzed without target identification. Amplification products were subjected to Illumina MiSeq sequencing, and our computational pipeline was used to distinguish intact and defective sequences as described before (5, 32). Monogram Biosciences performed neutralization assays for monoclonal antibodies (mAbs; PhenoSense mAb) on plasma collected at time of virologic failure and env amplicons from full-length intact proviruses at baseline (usually from birth). We defined full susceptibility to each bNAb as 90% inhibitory

concentration (IC\_{90}) of  $\leq$ 1.0 µg/ml and maximum percent inhibition of  $\geq$ 98% (3).

#### Study products and dosing

10-1074 was manufactured by MassBio under contract to the National Institute of Allergy and Infectious Diseases (NIAID) and dosed at 30 mg/kg iv every 4 weeks. VRC01LS was manufactured at the Vaccine Research Center, NIAID, and dosed at 30 mg/kg intravenously (iv) at entry and then continued at 15 mg/kg iv every 4 weeks. Administration of bNAbs occurred over approximately 60 min each. 10-1074 was given first, followed by VRC01LS, with a gap of approximately 10 to 15 min between infusions. Post-infusion monitoring for 2 to 4 hours occurred after the first infusion and 1 hour with subsequent infusions.

#### Prespecified objectives and definitions

Prespecified primary outcomes were adverse events through study end graded by NIAID Division of AIDS criteria (25) and the proportion of children maintaining HIV-1 RNA in plasma of <400 copies/ml through week 24 of the bNAb-only step. The protocol prespecified that  $\geq$ 30% of children maintaining HIV-1 RNA of <400 copies/ml for 24 weeks would likely represent a true effect of dual bNAbs on viral suppression. This percentage excluded overlap in 95% CIs with the CHER Study (14, 15), where the estimated probability of HIV-1 RNA suppression of <400 copies/ml at 6 months was 6% (range: 3 to 10%) after treatment interruption among children who started ART at younger than 12 weeks of age.

#### **Statistical analysis**

All raw, individual-level data for experiments where n < 20 are presented in data file S1. The sample size of children eligible for the EIT study allowed reasonable precision to estimate the proportion able to maintain HIV-1 RNA suppression of <400 copies/ml through week 24 of the bNAb-only step. Study objectives were analyzed under a proof-of-concept framework, without a control arm. Cumulative incidence of detectable viremia of ≥400 copies/ml during the bNAb-only step was estimated by Kaplan-Meier method. Characteristics of participants with ongoing viral suppression or viremia of ≥400 copies/ml were compared using Wilcoxon rank sum (for continuous variables) or Fisher's exact (for categorical variables) tests. Reported two-sided *P* values and CIs are presented as nominal, with a significance level set at 0.05. Analyses were conducted in SAS version 9.4.

#### **Supplementary Materials**

This PDF file includes: Figs. S1 to S3 Tables S1 and S2

Other Supplementary Material for this manuscript includes the following: Data file S1 MDAR Reproducibility Checklist

View/request a protocol for this paper from Bio-protocol.

#### **REFERENCES AND NOTES**

 M. Caskey, F. Klein, J. C. Lorenzi, M. S. Seaman, A. P. West Jr., N. Buckley, G. Kremer, L. Nogueira, M. Braunschweig, J. F. Scheid, J. A. Horwitz, I. Shimeliovich, S. Ben-Avraham, M. Witmer-Pack, M. Platten, C. Lehmann, L. A. Burke, T. Hawthorne, R. J. Gorelick, B. D. Walker, T. Keler, R. M. Gulick, G. Fatkenheuer, S. J. Schlesinger, M. C. Nussenzweig, Viraemia suppressed in HIV-1-infected humans by broadly neutralizing antibody 3BNC117. *Nature* **522**, 487–491 (2015).

- M. Caskey, T. Schoofs, H. Gruell, A. Settler, T. Karagounis, E. F. Kreider, B. Murrell, N. Pfeifer, L. Nogueira, T. Y. Oliveira, G. H. Learn, Y. Z. Cohen, C. Lehmann, D. Gillor, I. Shimeliovich, C. Unson-O'Brien, D. Weiland, A. Robles, T. Kummerle, C. Wyen, R. Levin, M. Witmer-Pack, K. Eren, C. Ignacio, S. Kiss, A. P. West Jr., H. Mouquet, B. S. Zingman, R. M. Gulick, T. Keler, P. J. Bjorkman, M. S. Seaman, B. H. Hahn, G. Fatkenheuer, S. J. Schlesinger, M. C. Nussenzweig, F. Klein, Antibody 10-1074 suppresses viremia in HIV-1-infected individuals. *Nat. Med.* 23, 185–191 (2017).
- C. Gaebler, L. Nogueira, E. Stoffel, T. Y. Oliveira, G. Breton, K. G. Millard, M. Turroja, A. Butler, V. Ramos, M. S. Seaman, J. D. Reeves, C. J. Petroupoulos, I. Shimeliovich, A. Gazumyan, C. S. Jiang, N. Jilg, J. F. Scheid, R. Gandhi, B. D. Walker, M. C. Sneller, A. Fauci, T. W. Chun, M. Caskey, M. C. Nussenzweig, Prolonged viral suppression with anti-HIV-1 antibody therapy. *Nature* 606, 368–374 (2022).
- T. Schoofs, F. Klein, M. Braunschweig, E. F. Kreider, A. Feldmann, L. Nogueira, T. Oliveira, J. C. Lorenzi, E. H. Parrish, G. H. Learn, A. P. West Jr., P. J. Bjorkman, S. J. Schlesinger, M. S. Seaman, J. Czartoski, M. J. McElrath, N. Pfeifer, B. H. Hahn, M. Caskey, M. C. Nussenzweig, HIV-1 therapy with monoclonal antibody 3BNC117 elicits host immune responses against HIV-1. *Science* 352, 997–1001 (2016).
- P. Garcia-Broncano, S. Maddali, K. B. Einkauf, C. Jiang, C. Gao, J. Chevalier, F. Z. Chowdhury, K. Maswabi, G. Ajibola, S. Moyo, T. Mohammed, T. Ncube, J. Makhema, P. Jean-Philippe, X. G. Yu, K. M. Powis, S. Lockman, D. R. Kuritzkes, R. Shapiro, M. Lichterfeld, Early antiretroviral therapy in neonates with HIV-1 infection restricts viral reservoir size and induces a distinct innate immune profile. *Sci. Transl. Med.* **11**, eaax7350 (2019).
- P. Palma, P. Zangari, C. Alteri, H. K. Tchidjou, E. C. Manno, G. Liuzzi, C. F. Perno, P. Rossi, A. Bertoli, S. Bernardi, Early antiretroviral treatment (eART) limits viral diversity over time in a long-term HIV viral suppressed perinatally infected child. *BMC Infect. Dis.* 16, 742 (2016).
- M. C. Sneller, J. Blazkova, J. S. Justement, V. Shi, B. D. Kennedy, K. Gittens, J. Tolstenko, G. McCormack, E. J. Whitehead, R. F. Schneck, M. A. Proschan, E. Benko, C. Kovacs, C. Oguz, M. S. Seaman, M. Caskey, M. C. Nussenzweig, A. S. Fauci, S. Moir, T. W. Chun, Combination anti-HIV antibodies provide sustained virological suppression. *Nature* 606, 375–381 (2022).
- C. K. Cunningham, E. J. McFarland, R. L. Morrison, E. V. Capparelli, J. T. Safrit, L. M. Mofenson, B. Mathieson, M. E. Valentine, C. Perlowski, B. Smith, R. Hazra, L. Purdue, P. Muresan, P. A. Harding, T. Mbengeranwa, L. G. Robinson, A. Wiznia, G. Theron, B. Lin, R. T. Bailer, J. R. Mascola, B. S. Graham; IMPAACT P1112 team, Safety, tolerability, and pharmacokinetics of the broadly neutralizing human immunodeficiency virus (HIV)-1 monoclonal antibody VRC01 in HIV-exposed newborn infants. J Infect Dis 23, 628–636 (2020).
- E. J. McFarland, C. K. Cunningham, P. Muresan, E. V. Capparelli, C. Perlowski, P. Morgan, B. Smith, R. Hazra, L. Purdue, P. A. Harding, G. Theron, H. Mujuru, A. Agwu, M. Purswani, M. H. Rathore, B. Flach, A. Taylor, B. C. Lin, A. B. McDermott, J. R. Mascola, B. S. Graham; International Maternal Pediatric Adolescent AIDS Clinical Trials Network (IMPAACT) P1112 Team, Safety, tolerability, and pharmacokinetics of a long-acting broadly neutralizing human immunodeficiency virus type 1 (HIV-1) monoclonal antibody VRC01LS in HIV-1exposed newborn infants. *J Infect Dis* **224**, 1916–1924 (2021).
- K. Maswabi, G. Ajibola, K. Bennett, E. V. Capparelli, P. Jean-Philippe, S. Moyo, T. Mohammed, O. Batlang, M. Sakoi, S. Lockman, J. Makhema, M. Lichterfeld, D. R. Kuritzkes, M. D. Hughes, R. L. Shapiro, Safety and efficacy of starting antiretroviral therapy in the first week of life. *Clin. Infect. Dis.* **72**, 388–393 (2021).
- P. Mendoza, H. Gruell, L. Nogueira, J. A. Pai, A. L. Butler, K. Millard, C. Lehmann, I. Suarez, T. Y. Oliveira, J. C. C. Lorenzi, Y. Z. Cohen, C. Wyen, T. Kummerle, T. Karagounis, C. L. Lu, L. Handl, C. Unson-O'Brien, R. Patel, C. Ruping, M. Schlotz, M. Witmer-Pack, I. Shimeliovich, G. Kremer, E. Thomas, K. E. Seaton, J. Horowitz, A. P. West Jr., P. J. Bjorkman, G. D. Tomaras, R. M. Gulick, N. Pfeifer, G. Fatkenheuer, M. S. Seaman, F. Klein, M. Caskey, M. C. Nussenzweig, Combination therapy with anti-HIV-1 antibodies maintains viral suppression. *Nature* 561, 479–484 (2018).
- K. Wagh, T. Bhattacharya, C. Williamson, A. Robles, M. Bayne, J. Garrity, M. Rist, C. Rademeyer, H. Yoon, A. Lapedes, H. Gao, K. Greene, M. K. Louder, R. Kong, S. A. Karim, D. R. Burton, D. H. Barouch, M. C. Nussenzweig, J. R. Mascola, L. Morris, D. C. Montefiori, B. Korber, M. S. Seaman, Optimal combinations of broadly neutralizing antibodies for prevention and treatment of HIV-1 Clade C infection. *PLOS Pathog.* 12, e1005520 (2016).
- A. Niesar, X. Lian, R. Hua, G. Ajibola, M. Pretorius Holme, K. Maswabi, S. Moyo, N. C. Maphorisa, T. Mohammed, M. Mosetlhi, K. M. Powis, D. Kuritzkes, R. L. Shapiro, M. Lichterfeld. "Viral reservoir landscape of children with HIV in Botswana treated with dual bNAbs," in *Proceedings of the 30th Conference on Retroviruses and Opportunistic Infections* (CROI, Seattle, 19 to 22 February 2023), Abstract 141.
- M. F. Cotton, A. Violari, K. Otwombe, R. Panchia, E. Dobbels, H. Rabie, D. Josipovic, A. Liberty, E. Lazarus, S. Innes, A. J. van Rensburg, W. Pelser, H. Truter, S. A. Madhi, E. Handelsman, P. Jean-Philippe, J. A. McIntyre, D. M. Gibb, A. G. Babiker; CHER Study Team, Early time-limited antiretroviral therapy versus deferred therapy in South African infants infected with HIV:

Results from the children with HIV early antiretroviral (CHER) randomised trial. Lancet **382**, 1555–1563 (2013).

- A. Violari, M. Chan, K. N. Otwombe, R. Panchia, P. Jean-Philippe, D. Gibb, M. Cotton, A. Babiker, "Time to viral rebound after stopping ART in children treated from infancy in CHER," in *Proceedings of the Conference on Retroviruses and Opportunistic Infections* (CROI, Boston, 4 to 7 March 2018), Abstract 137.
- S. F. Awan, M. Happe, A. R. Hofstetter, L. Gama, Broadly neutralizing antibodies for treatment and prevention of HIV-1 infection. *Curr. Opin. HIV AIDS* 17, 247–257 (2022).
- B. Noailly, M. Yaugel-Novoa, J. Werquin, F. Jospin, D. Drocourt, T. Bourlet, N. Rochereau, S. Paul, Antiviral activities of HIV-1-specific human broadly neutralizing antibodies are isotype-dependent. *Vaccines (Basel)* **10**, 903 (2022).
- C. L. Lu, D. K. Murakowski, S. Bournazos, T. Schoofs, D. Sarkar, A. Halper-Stromberg, J. A. Horwitz, L. Nogueira, J. Golijanin, A. Gazumyan, J. V. Ravetch, M. Caskey, A. K. Chakraborty, M. C. Nussenzweig, Enhanced clearance of HIV-1-infected cells by broadly neutralizing antibodies against HIV-1 in vivo. *Science* 352, 1001–1004 (2016).
- R. M. Lynch, E. Boritz, E. E. Coates, A. DeZure, P. Madden, P. Costner, M. E. Enama, S. Plummer, L. Holman, C. S. Hendel, I. Gordon, J. Casazza, M. Conan-Cibotti, S. A. Migueles, R. Tressler, R. T. Bailer, A. McDermott, S. Narpala, S. O'Dell, G. Wolf, J. D. Lifson, B. A. Freemire, R. J. Gorelick, J. P. Pandey, S. Mohan, N. Chomont, R. Fromentin, T. W. Chun, A. S. Fauci, R. M. Schwartz, R. A. Koup, D. C. Douek, Z. Hu, E. Capparelli, B. S. Graham, J. R. Mascola, J. E. Ledgerwood; VRC 601 Study Team, Virologic effects of broadly neutralizing antibody VRC01 administration during chronic HIV-1 infection. *Sci. Transl. Med.* 7, 319ra206 (2015).
- K. J. Bar, M. C. Sneller, L. J. Harrison, J. S. Justement, E. T. Overton, M. E. Petrone, D. B. Salantes, C. A. Seamon, B. Scheinfeld, R. W. Kwan, G. H. Learn, M. A. Proschan, E. F. Kreider, J. Blazkova, M. Bardsley, E. W. Refsland, M. Messer, K. E. Clarridge, N. B. Tustin, P. J. Madden, K. Oden, S. J. O'Dell, B. Jarocki, A. R. Shiakolas, R. L. Tressler, N. A. Doria-Rose, R. T. Bailer, J. E. Ledgerwood, E. V. Capparelli, R. M. Lynch, B. S. Graham, S. Moir, R. A. Koup, J. R. Mascola, J. A. Hoxie, A. S. Fauci, P. Tebas, T. W. Chun, Effect of HIV antibody VRC01 on viral rebound after treatment interruption. *N. Engl. J. Med.* **375**, 2037–2050 (2016).
- J. E. Ledgerwood, E. E. Coates, G. Yamshchikov, J. G. Saunders, L. Holman, M. E. Enama, A. DeZure, R. M. Lynch, I. Gordon, S. Plummer, C. S. Hendel, A. Pegu, M. Conan-Cibotti, S. Sitar, R. T. Bailer, S. Narpala, A. McDermott, M. Louder, S. O'Dell, S. Mohan, J. P. Pandey, R. M. Schwartz, Z. Hu, R. A. Koup, E. Capparelli, J. R. Mascola, B. S. Graham; VRC 602 Study Team, Safety, pharmacokinetics and neutralization of the broadly neutralizing HIV-1 human monoclonal antibody VRC01 in healthy adults. *Clin. Exp. Immunol.* **182**, 289–301 (2015).
- L. Corey, P. B. Gilbert, M. Juraska, D. C. Montefiori, L. Morris, S. T. Karuna, S. Edupuganti, N. M. Mgodi, A. C. deCamp, E. Rudnicki, Y. Huang, P. Gonzales, R. Cabello, C. Orrell, J. R. Lama, F. Laher, E. M. Lazarus, J. Sanchez, I. Frank, J. Hinojosa, M. E. Sobieszczyk, K. E. Marshall, P. G. Mukwekwerere, J. Makhema, L. R. Baden, J. I. Mullins, C. Williamson, J. Hural, M. J. McElrath, C. Bentley, S. Takuva, M. M. Gomez Lorenzo, D. N. Burns, N. Espy, A. K. Randhawa, N. Kochar, E. Piwowar-Manning, D. J. Donnell, N. Sista, P. Andrew, J. G. Kublin, G. Gray, J. E. Ledgerwood, J. R. Mascola, M. S. Cohen; HVTN 704/HPTN 085; HVTN 703/HPTN 081 Study Teams, Two randomized trials of neutralizing antibodies to prevent HIV-1 acquisition. N. Engl. J. Med. 384, 1003–1014 (2021).
- M. Mosetlhi, G. Ajibola, R. Haghighat, O. Batlang, K. Maswabi, M. Pretorius Holme, K. M. Powis, S. Lockman, J. Makhema, M. Lichterfeld, D. Kuritzkes, R. L. Shapiro, "Caregivers of children with HIV in Botswana prefer monthly IV bNAbs to daily oral ART," in Proceedings of the 30th Conference on Retroviruses and Opportunistic Infections (CROI, Seattle, 19 to 22 February 2023), abstract 828.
- E. V. Capparelli, G. Ajibola, K. Maswabi, M. P. Holme, K. Bennett, K. M. Powis, S. Moyo, T. Mohammed, C. Maphorisa, M. D. Hughes, K. E. Seaton, G. D. Tomaras, S. Mosher, A. Taylor, S. O'Connell, S. Narpala, A. McDermott, M. Caskey, L. Gama, S. Lockman, P. Jean-Philippe, J. Makhema, D. R. Kuritzkes, M. Lichterfeld, R. L. Shapiro; Tatelo Study Team, Safety and pharmacokinetics of intravenous 10-1074 and VRC01LS in young children. J. Acquir. Immune Defic. Syndr. 91, 182–188 (2022).
- Division of AIDS Table for Grading the Severity of Adult and Pediatric Adverse Events, version 2.1 July 2017; https://rsc.niaid.nih.gov/sites/default/files/daidsgradingcorrectedv21.pdf [accessed 8 July 2021].
- M. R. Gaudinski, E. E. Coates, K. V. Houser, G. L. Chen, G. Yamshchikov, J. G. Saunders, L. A. Holman, I. Gordon, S. Plummer, C. S. Hendel, M. Conan-Cibotti, M. G. Lorenzo, S. Sitar, K. Carlton, C. Laurencot, R. T. Bailer, S. Narpala, A. B. McDermott, A. M. Namboodiri, J. P. Pandey, R. M. Schwartz, Z. Hu, R. A. Koup, E. Capparelli, B. S. Graham, J. R. Mascola, J. E. Ledgerwood; VRC 606 Study Team, Safety and pharmacokinetics of the Fc-modified HIV-1 human monoclonal antibody VRC01LS: A phase 1 open-label clinical trial in healthy adults. *PLoS Med.* **15**, e1002493 (2018).
- Y. Z. Cohen, A. L. Butler, K. Millard, M. Witmer-Pack, R. Levin, C. Unson-O'Brien, R. Patel, I. Shimeliovich, J. C. C. Lorenzi, J. Horowitz, S. R. Walsh, S. Lin, J. A. Weiner, A. Tse, A. Sato, C. Bennett, B. Mayer, K. E. Seaton, N. L. Yates, L. R. Baden, A. C. deCamp, M. E. Ackerman, M. S.

Seaman, G. D. Tomaras, M. C. Nussenzweig, M. Caskey, Safety, pharmacokinetics, and immunogenicity of the combination of the broadly neutralizing anti-HIV-1 antibodies 3BNC117 and 10-1074 in healthy adults: A randomized, phase 1 study. *PLOS ONE* **14**, e0219142 (2019).

- P. Bharadwaj, C. Riekofski, S. Lin, M. S. Seaman, D. A. Garber, D. Montefiori, M. Sarzotti-Kelsoe, M. E. Ackerman, J. A. Weiner, Implementation of a three-tiered approach to identify and characterize anti-drug antibodies raised against HIV-specific broadly neutralizing antibodies. J. Immunol. Methods 479, 112764 (2020).
- C. Jiang, X. Lian, C. Gao, X. Sun, K. B. Einkauf, J. M. Chevalier, S. M. Y. Chen, S. Hua, B. Rhee, K. Chang, J. E. Blackmer, M. Osborn, M. J. Peluso, R. Hoh, M. Somsouk, J. Milush, L. N. Bertagnolli, S. E. Sweet, J. A. Varriale, P. D. Burbelo, T. W. Chun, G. M. Laird, E. Serrao, A. N. Engelman, M. Carrington, R. F. Siliciano, J. M. Siliciano, S. G. Deeks, B. D. Walker, M. Lichterfeld, X. G. Yu, Distinct viral reservoirs in individuals with spontaneous control of HIV-1. *Nature* 585, 261–267 (2020).
- G. Q. Lee, N. Orlova-Fink, K. Einkauf, F. Z. Chowdhury, X. Sun, S. Harrington, H. H. Kuo, S. Hua, H. R. Chen, Z. Ouyang, K. Reddy, K. Dong, T. Ndung'u, B. D. Walker, E. S. Rosenberg, X. G. Yu, M. Lichterfeld, Clonal expansion of genome-intact HIV-1 in functionally polarized Th1 CD4<sup>+</sup> T cells. J. Clin. Invest. **127**, 2689–2696 (2017).
- M. J. Buzon, H. Sun, C. Li, A. Shaw, K. Seiss, Z. Ouyang, E. Martin-Gayo, J. Leng, T. J. Henrich, J. Z. Li, F. Pereyra, R. Zurakowski, B. D. Walker, E. S. Rosenberg, X. G. Yu, M. Lichterfeld, HIV-1 persistence in CD4+ T cells with stem cell-like properties. *Nat. Med.* 20, 139–142 (2014).
- C. A. Hartana, P. Garcia-Broncano, Y. Rassadkina, X. Lian, C. Jiang, K. B. Einkauf, K. Maswabi, G. Ajibola, S. Moyo, T. Mohammed, C. Maphorisa, J. Makhema, Y. Yuki, M. Martin, K. Bennett, P. Jean-Philippe, M. Viard, M. D. Hughes, K. M. Powis, M. Carrington, S. Lockman, C. Gao, X. G. Yu, D. R. Kuritzkes, R. Shapiro, M. Lichterfeld, Immune correlates of HIV-1 reservoir cell decline in early-treated infants. *Cell Rep.* **40**, 111126 (2022).

Acknowledgments: We thank the Tatelo Study participants and their families. We thank the Tatelo and EIT Study teams and collaborators, including D. Babuile, R. Bowman, C. Brackett, L. Bunhu, A. Carnacchi, L. Colson, T. Cordwell, J. Disaro, L. Esele, T. Frank, O. Kgakge, T. Kebopetswe, N. Kelentse, B. Lin, J. Lucas, K. Lvons, A. Maigwa, R. Madison, P. Mapenshi, M. Matshaba, S. Masopa, S. McMillan, C. Mdluli, L. Melton, M. Mmalane, M. Mosetlhi, A. Motlhanka, S. Mudrak, S. Narpala, M. Nagvi, T. Ncube, S. Ndongwe, M. Ngwaca, M. Oabona, S. Othusitse, G. Pelontle, O. Pule, L. Purdue, C. Reding, M. Sarzotti-Kelsoe, T. Sekoto, N. Seonyatseng, D. Tumagole, and J. Weiner; and, at Labcorp-Monogram Biosciences, C. Petropoulos, K. Strommen, Y. Lie, and T. Persyn as well as the Clinical Reference Lab. We also thank the Botswana Ministry of Health and Wellness and the Tatelo Safety Monitoring Committee, including G. John-Stewart, K. Luzuriaga, L. Mazhani, P. Tebas, T. Fenton, and J. Lindsev. Funding: This work was funded by the U.S. National Institute of Allergy and Infectious Diseases cooperative agreement U01 AI135940 (to R.L.S., D.R.K., and M.L.), R.L.S. and M.L. are members of the PAVE Martin Delaney Collaboratory for HIV Cure Research (UM1 AI164566). M.L. is a member of the EPIICAL consortium, funded by ViiV Healthcare. S.M. was supported by the U.S. NIH Fogarty International Center (K43 TW012350). The contents of this article are solely the responsibility of the authors and do not necessarily represent the official positions of the funding agencies. Author contributions: R.L.S., M.H., S.L., J.M., D.R.K., M.L., and E.V.C. conceptualized the study. R.L.S., D.R.K., M.L., P.J.-P., D.E.Y., M. Caskey, and L.G. acquired the study drugs. G.A., K.M., K.M.P., M.S., O.B., S.M., T.M., C.M., A.N., J.D.R., M.A.R., M.E.A., A.M., and M. Cooper collected or generated the data, M.H., B.S.N., K.B., R.L.S., G.A., Z.H., F.G., J.D.R., M.A.R., C.G., X.Y., and E.V.C. analyzed the data. R.L.S., G.A., M.P.H., E.V.C., M.H., B.S.N., D.R.K., and M.L. wrote the original draft of the manuscript. All authors reviewed and edited the manuscript. Competing interests: R.L.S., M.H., S.L., and J.M. serve on the governing boards of the Botswana Harvard Health Partnership. K.B. consulted with Harvard T. H. Chan School of Public Health. Massachusetts Eve and Ear Infirmary (MEEI), and the University of Alabama at Birmingham. J.D.R. is an employee and stockholder of Labcorp-Monogram Biosciences. D.E.Y. was formerly an unpaid technical advisor for the nonprofit organizations Cover The Globe and Maipelo Trust. D.R.K. has served as a consultant for AbbVie, Gilead Sciences, GlaxoSmithKline, Janssen Pharmaceuticals, Merck, and ViiV Healthcare; has received research support from Gilead Sciences, Merck, and ViiV Healthcare; has received speaking honoraria from Gilead Sciences and Janssen Pharmaceuticals; and has provided expert testimony for Gilead. All other authors declare that they have no competing interests. Data and materials availability: All data associated with this study are present in the paper or the Supplementary Materials. Deidentified or partially deidentified data, as appropriate, will be made available after the completion of the study to researchers with an approved protocol who complete a data use agreement. All inquiries should be sent to the corresponding author.

Submitted 3 February 2023 Accepted 26 April 2023 Published 5 July 2023 10.1126/scitranslmed.adh0004

# **Science** Translational Medicine

## Broadly neutralizing antibody treatment maintained HIV suppression in children with favorable reservoir characteristics in Botswana

Roger L. Shapiro, Gbolahan Ajibola, Kenneth Maswabi, Michael Hughes, Bryan S. Nelson, Aischa Niesar, Molly Pretorius Holme, Kathleen M. Powis, Maureen Sakoi, Oganne Batlang, Sikhulile Moyo, Terence Mohammed, Comfort Maphorisa, Kara Bennett, Zixin Hu, Francoise Giguel, Jacqueline D. Reeves, Michael A. Reeves, Ce Gao, Xu Yu, Margaret E. Ackerman, Adrian McDermott, Marlene Cooper, Marina Caskey, Lucio Gama, Patrick Jean-Philippe, Dwight E. Yin, Edmund V. Capparelli, Shahin Lockman, Joseph Makhema, Daniel R. Kuritzkes, and Mathias Lichterfeld

*Sci. Transl. Med.*, **15** (703), eadh0004. DOI: 10.1126/scitransImed.adh0004

View the article online https://www.science.org/doi/10.1126/scitransImed.adh0004 Permissions https://www.science.org/help/reprints-and-permissions

Use of this article is subject to the Terms of service

Science Translational Medicine (ISSN) is published by the American Association for the Advancement of Science. 1200 New York Avenue NW, Washington, DC 20005. The title Science Translational Medicine is a registered trademark of AAAS. Copyright © 2023 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works